

Tetrahedron Letters, Vol. 35, No. 23, pp. 3845-3848, 1994 Elsevier Science Ltd Printed in Great Britain 0040-4039/94 \$7.00+0.00

0040-4039(94)E0696-U

A NOVEL INHIBITOR OF HUMAN α -L-FUCOSIDASE: ENANTIOSELECTIVE SYNTHESIS OF L-FUCOAMIDRAZONE

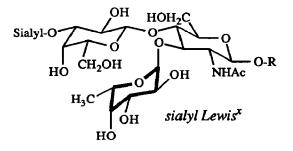
David J. A. Schedler[‡], Benjamin R. Bowen[§] and Bruce Ganem^{‡*}

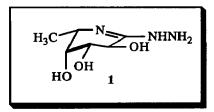
[‡]Department of Chemistry, Baker Laboratory Cornell University, Ithaca, NY 14853-1301 USA

Summit, NJ 07901 USA
§Pharmaceuticals Division, CIBA-GEIGY Corporation

Abstract: A short, chiral synthesis of the title inhibitor 1 is reported from D-galactosamine by chain end interchange. The L-fucoamidrazone is a good competitive inhibitor of human α -L-fucosidase (K_i = 820 nM).

The phenomenon of cell-cell adhesion is one of several key recognition events between leukocytes and endothelial cells responsible for orchestrating the appropriate immune response to inflammation.¹ Cytokines trigger endothelial cells to produce cell-surface endothelial-leukocyte adhesion molecules (ELAMs) which bind to a tetrasaccharide, sialyl Lewis^x, expressed on the leukocyte surface. Sialyl Lewis^x (shown below) is the minimal carbohydrate ligand for many ELAMs, and adhesion has been shown to depend critically on the presence of a 1,3linked L-fucose residue (bold ring in figure).²⁻⁴ For these reasons, inhibitors of the corresponding fucoseprocessing enzymes which mediate trimming (L-fucosidase) and attachment (L-fucosyltransferase) are currently of considerable interest, not only for mechanistic study, but also as prospective anti-inflammatory and antitumor drugs. Here we report the synthesis and bioevaluation of L-fucoamidrazone 1 which, as expected from studies of related monosaccharide analogs,⁵ is a potent inhibitor of human α -L-fucosidase.



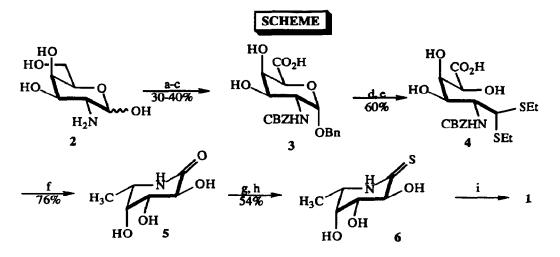


Many glycosidases are inhibited by polyhydroxylated piperidines and pyrrolidines resembling the enzyme's cognate saccharide substrate, prompting several groups to target fucose-related polyhydroxylated heterocycles as prospective fucosidase inhibitors. Thus Fleet's group⁶ and others⁷ have prepared 1,5-dideoxy-1,5-imino-L-fucitol, which is a potent competitive inhibitor of human α -L-fucosidase (K_i = 10⁻⁸ M). Several closely-related iminoalditols and glycals also inhibit fucosidase, some with K_i values in the low micromolar range.⁸ A complementary approach involved the synthesis of iminoalditol-containing Lewis^x and sialyl Lewis^x oligosaccharides, ⁹ as well as sulfur-linked disaccharides of fucose.¹⁰ No bioassays on the former were reported, and the latter substances required millimolar concentrations for fucosidase inhibition.

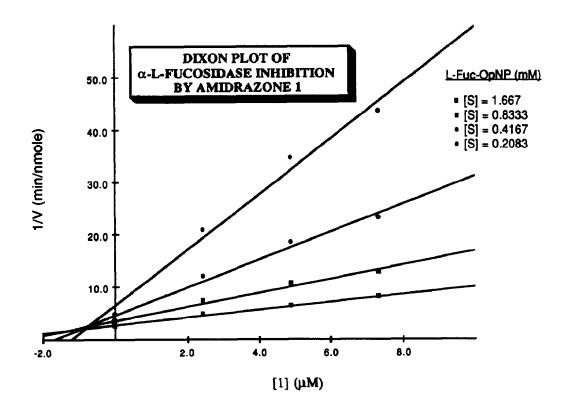
Amidrazone analogs of D-monosaccharides have been shown to be potent and effective inhibitors of a wide range of glycosidases.⁵ Based on that earlier methodology, a synthesis of L-fucoamidrazone 1 required access to the known¹¹ L-fuconic- δ -lactam 5, for which a more expedient synthesis has now been developed from D-galactosamine 2 using a chain end interchange strategy (Scheme). Thus, the known¹² galacturonic acid 3 was prepared by platinum and oxygen dehydrogenation of protected 2 in good yield on a multigram scale.¹³ After hydrolysis of the glycosidic bond in 3, thioacetalization of the latent aldehyde using ethanethiol afforded 4 in 60% overall yield. Upon exposure under acidic conditions to Raney nickel, desulfurization was accompanied with N-deprotection and concomitant cyclization to generate the known lactam 5 (76%), whose physical and spectroscopic properties were identical with published values.¹¹ Exhaustive silylation of 5, followed by treatment with Lawesson's reagent furnished thionolactam 6 after desilylation in the acidic workup. Reaction of 6 with hydrazine in CH₃OH gave the target amidrazone 1 in 90% yield after flash column chromatography.

The activity of 1 as a fucosidase inhibitor was evaluated using recombinant human α -L-fucosidase. Under steady-state assay conditions¹⁴ using p-nitrophenyl- α -L-fucopyranoside (Fuc-OpNP) as substrate (K_M = 230 μ M, citrate buffer, pH 5.0) the enzyme was well-behaved, and Dixon plots of 1/V versus [I] revealed clean competitive inhibition in the presence of L-fucose (K_i = 400 μ M). Competitive inhibition was also observed in the presence of 1 (see Figure; facing page), with K_i = 820 nM. Thus with its appropriately flattened (i.e. half-chair) conformation, ease of protonation, and good stability, L-fucoamidrazone 1 represents a promising new lead in the search for potent fucosidase inhibitors, including sialyl Lewis^x oligosaccharides incorporating 1 and its congeners.

ACKNOWLEDGMENT: We thank the National Institutes of Health (GM 35712) for generous financial assistance. Support of the Cornell NMR Facility by the NSF and NIH is gratefully acknowledged.



(a) CBZCl, H₂O; (b) BnOH, CH₃COCl; (c) Pt, O₂, H₂O-EtOH; (d) 1:4 TFA:H₂O, 90°C; (e) EtSH, HCl; (f) RaNi-EtOH; (g) TMSCl, (TMS)₂NH, pyr; (h) Lawesson's reagent, C₆H₆, reflux, 2 h; then CH₃OH-HCl; (i) NH₂NH₂-CH₃OH, 0°C, 3 h.



REFERENCES AND NOTES

- (a) Lasky, L. A. Science 1992, 258, 964; (b) Feizi, T. TIBS 1991, 16, 84; (c) Springer, T. Nature 1990, 346, 425.
- Lowe, J. B.; Stoolman, L. M.; Nair, R. P.; Larsen, R. D.; Berhend, T. L.; Marks, R. M. Cell 1990, 63, 475.
- Berg, E. L.; Robinson, M. K.; Mansson, O.; Butcher, E. C.; Magnani, J. L. J. Biol. Chem. 1991, 266, 14869.
- Tyrrell, D.; James, P.; Rao, N.; Foxall, C.; Abbas, S.; Dasgupta, F.; Nashed, M.; Hasegawa, A.; Kiso, M.; Asa, D.; Kidd, J.; Brandley, B. K. Proc. Natl. Acad. Sci. U.S.A. 1991, 88, 10372.
- 5. Papandreou, G.; Tong, M. K.; Ganem, B. J. Am. Chem. Soc. 1993, 115, 11682.
- (a) Fleet, G. W. J.; Shaw, A. N.; Evans, S. V.; Fellows, L. E. J. Chem. Soc. Chem. Commun. 1985, 841; (b) Winchester, B.; Barker, C.; Baines, S.; Jacob, G. S.; Namgoong, S. K.; Fleet, G. Biochem. J. 1990, 265, 277.
- (a) Paulsen, H.; Matzke, M. Liebigs Ann. Chem. 1988, 1121; (b) Takahashi, S.; Kuzuhara, H. Chem. Lett. 1992, 21.
- (a) Fleet, G. W. J.; Namgoong, S. K.; Barker, C.; Baines, S.; Jacob, G. S.; Winchester, B. Tetrahedron Lett. 1989, 30, 4439; (b) Hashimoto, H.; Hayakawa, M. Chem. Lett. 1989, 1881; (c) Wang, Y.-F.; Dumas, D. P.; Wong, C.-H. Tetrahedron Lett. 1993, 34, 403; (d) Dumas, D. P.; Kajimoto, T.; Liu, K. K.-C.; Wong, C.-H.; Berkowitz, D. B.; Danishefsky, S. J. Bioorg. Med. Chem. Lett. 1992, 2, 33.
- 9. Furui, H.; Kiso, M.; Hasegawa, A. Carbohydr. Res. 1992, 229, C1.
- 10. Hashimoto, H.; Shimada, K.; Horito, S. Tetrahedron Lett. 1993, 34, 4953.
- Fleet, G. W. J.; Ramsden, N. G.; Dwek, R. A.; Rademacher, T. W.; Fellows, L. E.; Nash, R. J.; Green,
 D. St.-C.; Winchester, B. J. Chem. Soc. Chem. Commun. 1988, 483.
- 12. Heyns, K.; Beck, M. Chem. Ber. 1957, 90, 2443.
- 13. Satisfactory ¹H, ¹³C-NMR, IR and mass spectrometric data were obtained for all new compounds.
- 14. Alhadeff, J. A.; Miller, A. L.; Wenger, D. A.; O'Brien, J. S. Clinica Chimica Acta 1974, 57, 307.

(Received in USA 1 March 1994; revised 7 April 1994; accepted 8 April 1994)